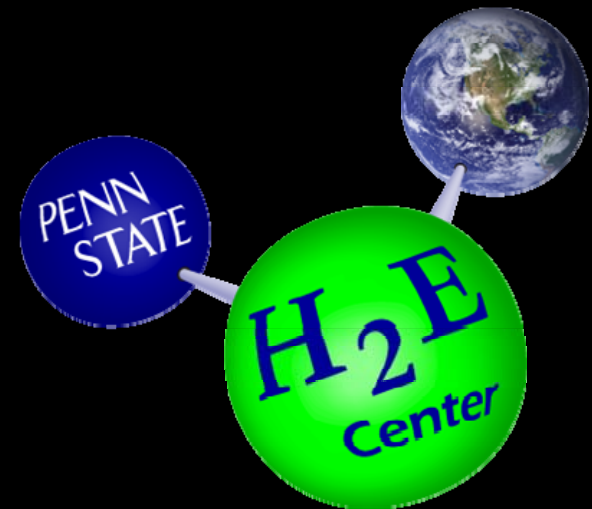


# Electricity and hydrogen production using microbial fuel cell-based technologies

*Bruce E. Logan*  
Penn State University

Engineering  
Environmental  
Institute

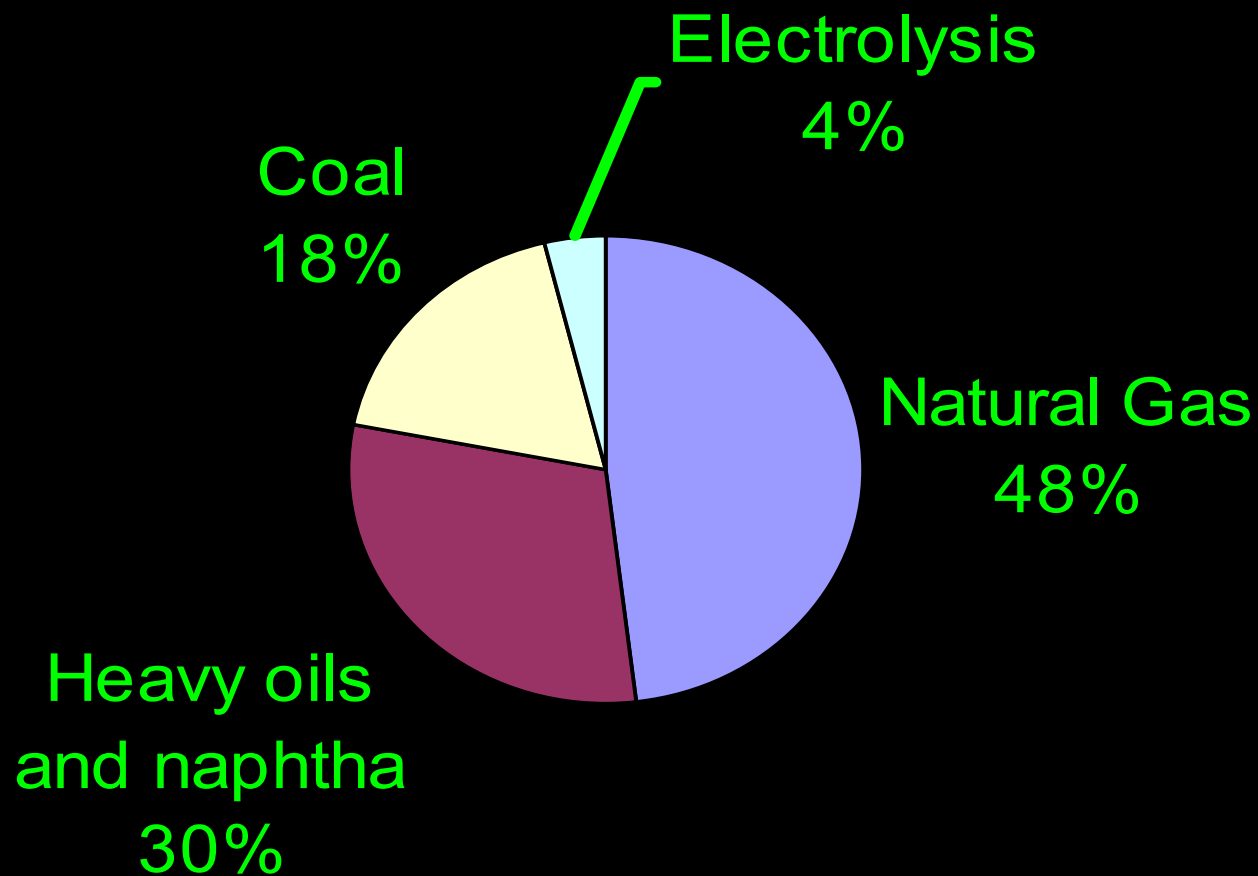


# Ethanol versus H<sub>2</sub> from Glucose

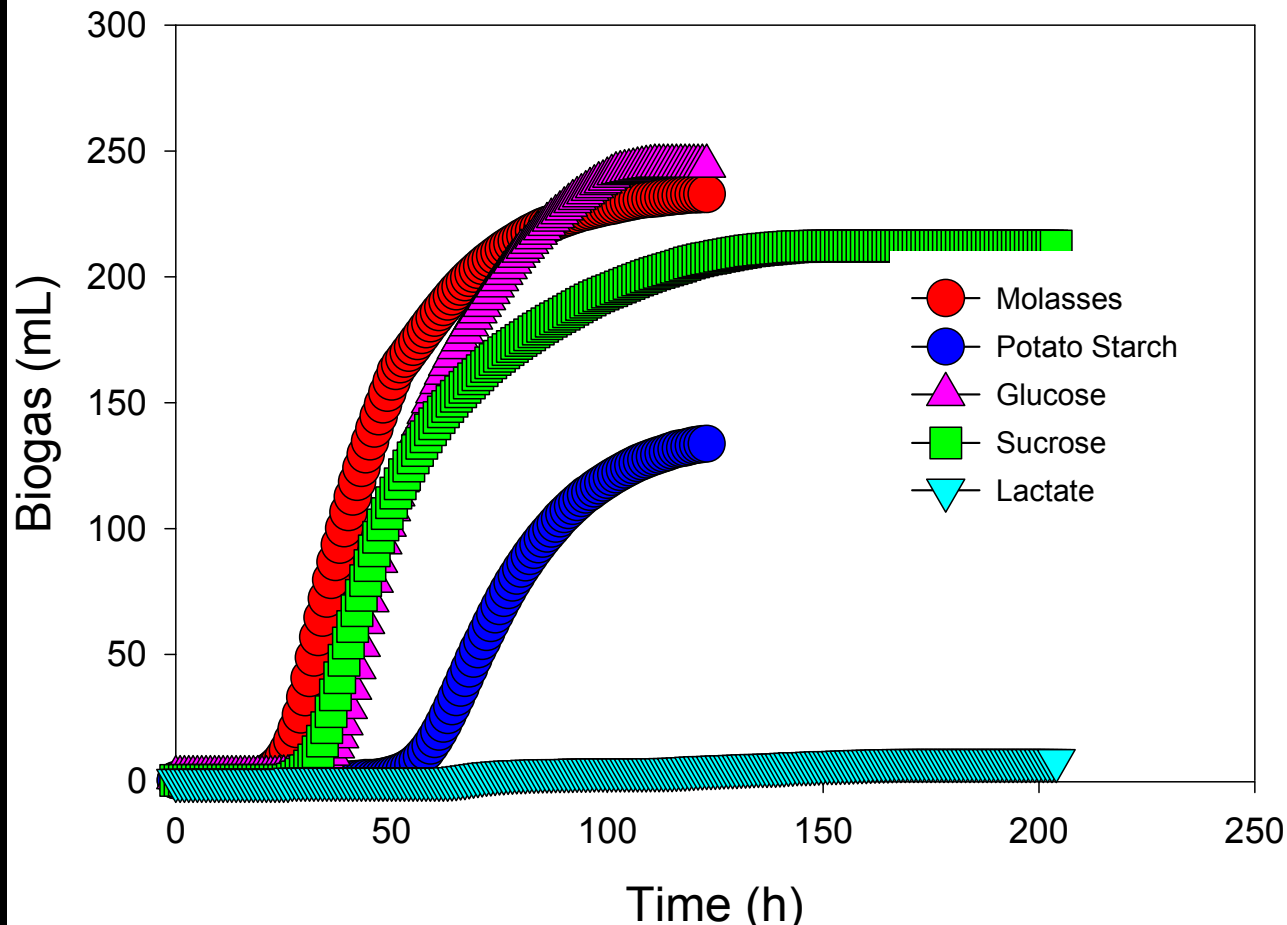
- Glucose:  $\Delta H_c = 2808 \text{ kJ/mol}$
- Ethanol
  - Produce 2 ethanol per glucose by fermentation
  - $2 \times \Delta H_c = 2 \times 1367 \text{ kJ/mol} \Rightarrow \Delta H_c = 2734 \text{ kJ/mol}$
- Hydrogen
  - Produce up to 12 H<sub>2</sub> per glucose
  - $12 \times \Delta H_c = 12 \times 286 \text{ kJ/mol} \Rightarrow \Delta H_c = 3430 \text{ kJ/mol}$
- DOE
  - 10-12 mol-H<sub>2</sub>/mol-glucose needed to make biological H<sub>2</sub> production feasible (9.6 mol H<sub>2</sub> =  $\Delta H_c$  for 2 ethanol from glucose)



# Current sources of H<sub>2</sub> Production



# H<sub>2</sub> can be biologically produced from bacterial fermentation of sugars



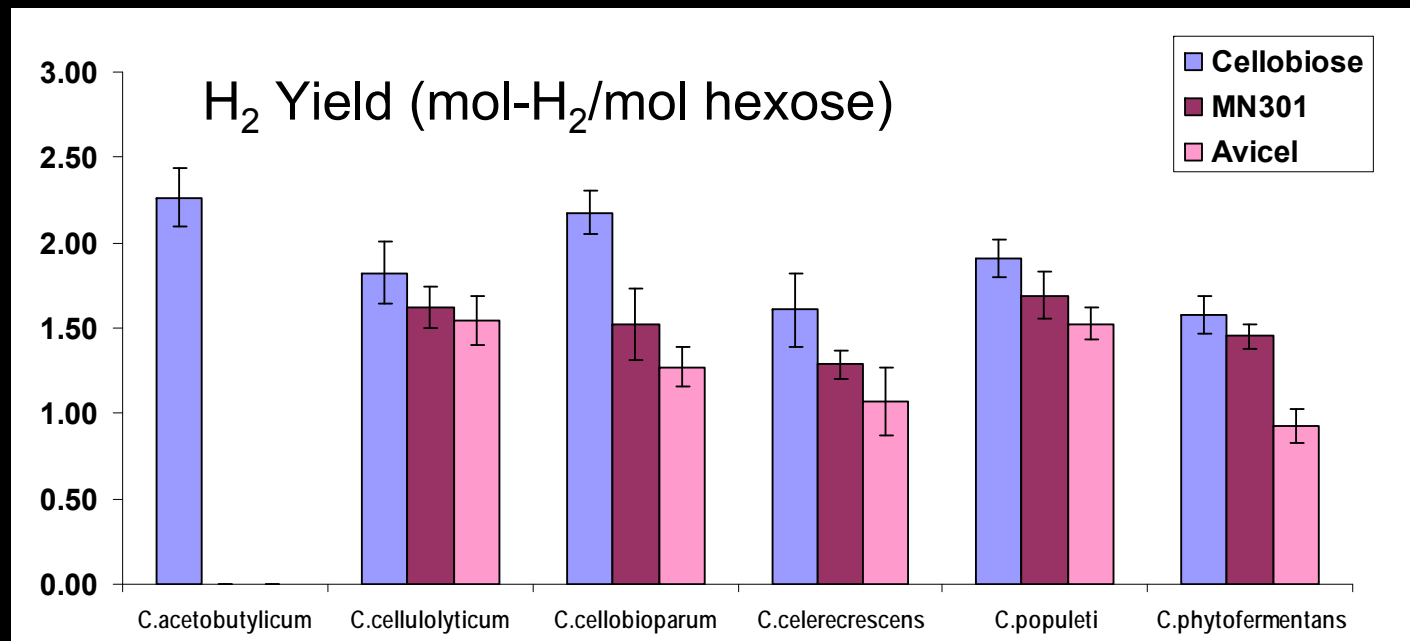
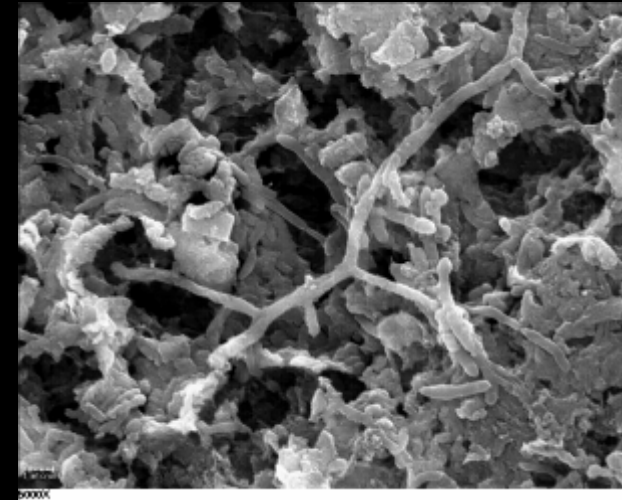
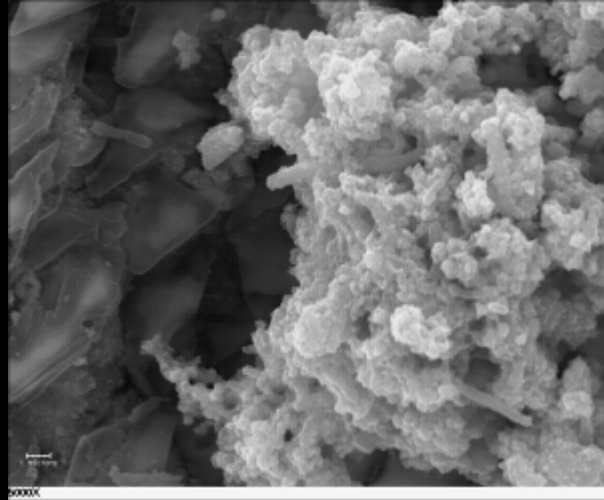
Biogas:

- 60% H<sub>2</sub>

- 40% CO<sub>2</sub>



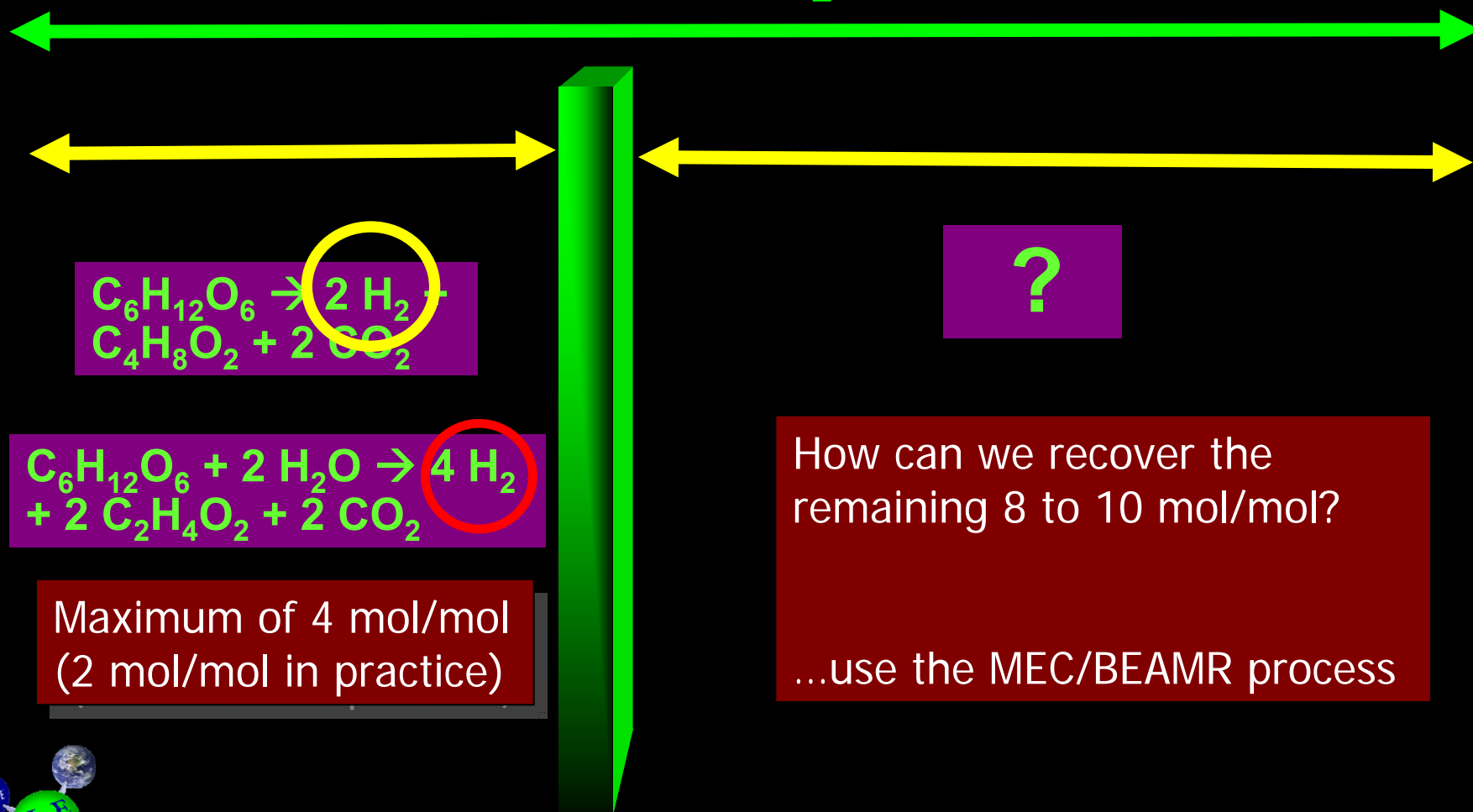
# H<sub>2</sub> can be produced by *cellulose fermentation*



Source: Ren et al., *J. Appl. Microbiol.* (2007)

# The “fermentation barrier” limits H<sub>2</sub> yields

Maximum: 12 mol-H<sub>2</sub>/mol-hexose



# Energy Production using MFC technologies

- Electricity production using microbial fuel cells (MFCs)
- H<sub>2</sub> Production from biomass using the MEC/BEAMR process: overcoming the “fermentation barrier”
- A path to renewable energy



# Demonstration of a Microbial Fuel Cell (MFC)

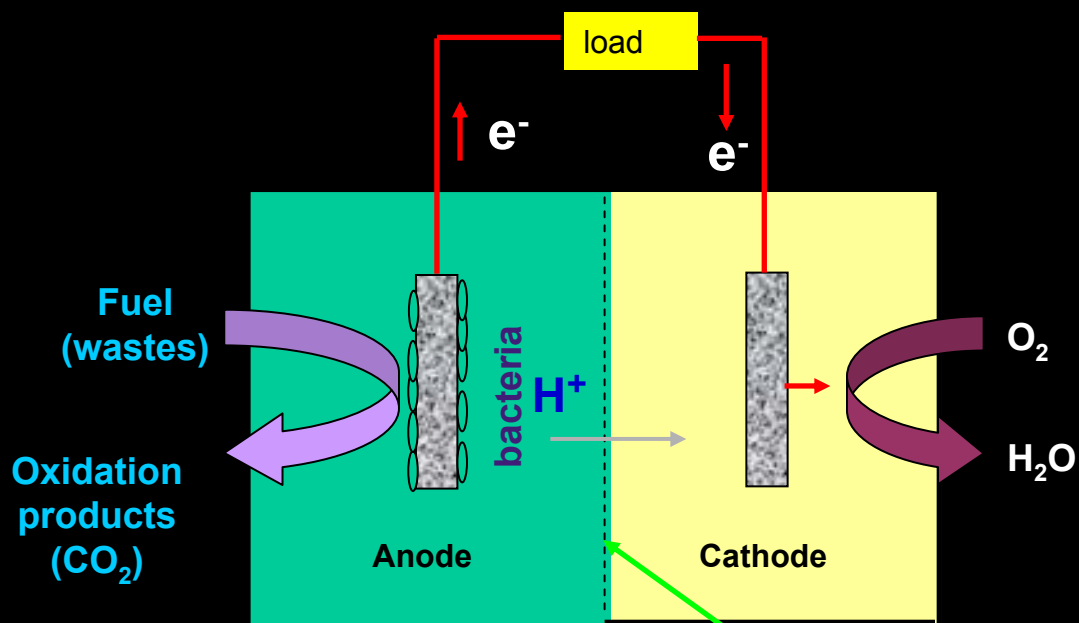


MFC webcam (live video of an MFC running a fan)

[www.engr.psu.edu/mfccam](http://www.engr.psu.edu/mfccam)



# Microbial Fuel Cells: Aqueous cathode



Proton exchange  
membrane (PEM)

- *Methanogenesis*

- Generation of methane
- Methanogens
- Anaerobic digesters

- *Electrogenesis*

- Generation of electricity
- Exoelectrogens
- Microbial fuel cells (MFCs)

- *Electrohydrogenesis*

- Generation of H<sub>2</sub> gas
- Exoelectrogens
- Microbial electrolysis cells (MECs)

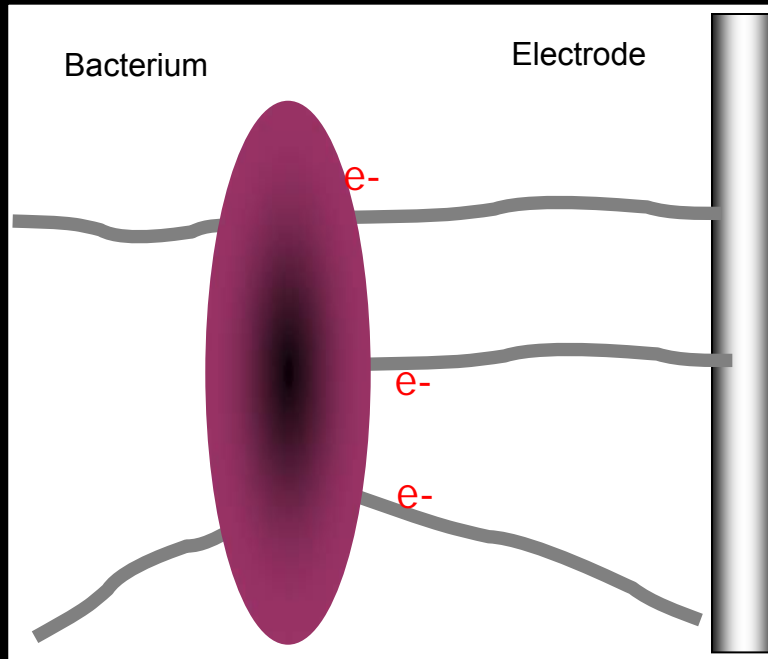


# Microbial community analysis (DGGE)

Inoculum (substrate)	Community	Reference
River sediment (glucose+glutamic acid)	$\alpha$ - <i>Proteobacteria</i> (mainly <i>Actinobacteria</i> )	Phung et al. (2004)
River sediment (river water)	$\beta$ - <i>Proteobacteria</i> (related to <i>Leptothrix</i> spp.)	Phung et al. (2004)
Marine sediment (cysteine)	$\gamma$ - <i>Proteobacteria</i> , 40% <i>Shewanella affinis</i> KMM, then <i>Vibrio</i> spp. and <i>Pseudoalteromonas</i> sp.	Logan et al. 2005
Wastewater (starch)	36%=unidentified, 25%= $\beta$ - and 20%= $\alpha$ - <i>Proteobacteria</i> , and 19%= <i>Cytophaga</i> + <i>Flexibacter</i> + <i>Bacterioides</i>	Kim et al. (2004)
Wastewater (acetate)	24%= $\alpha$ -, 7%= $\beta$ -, 21%= $\gamma$ - , 21%= $\delta$ - <i>Proteobacteria</i> ; 27%=others	Lee et al. (2003)

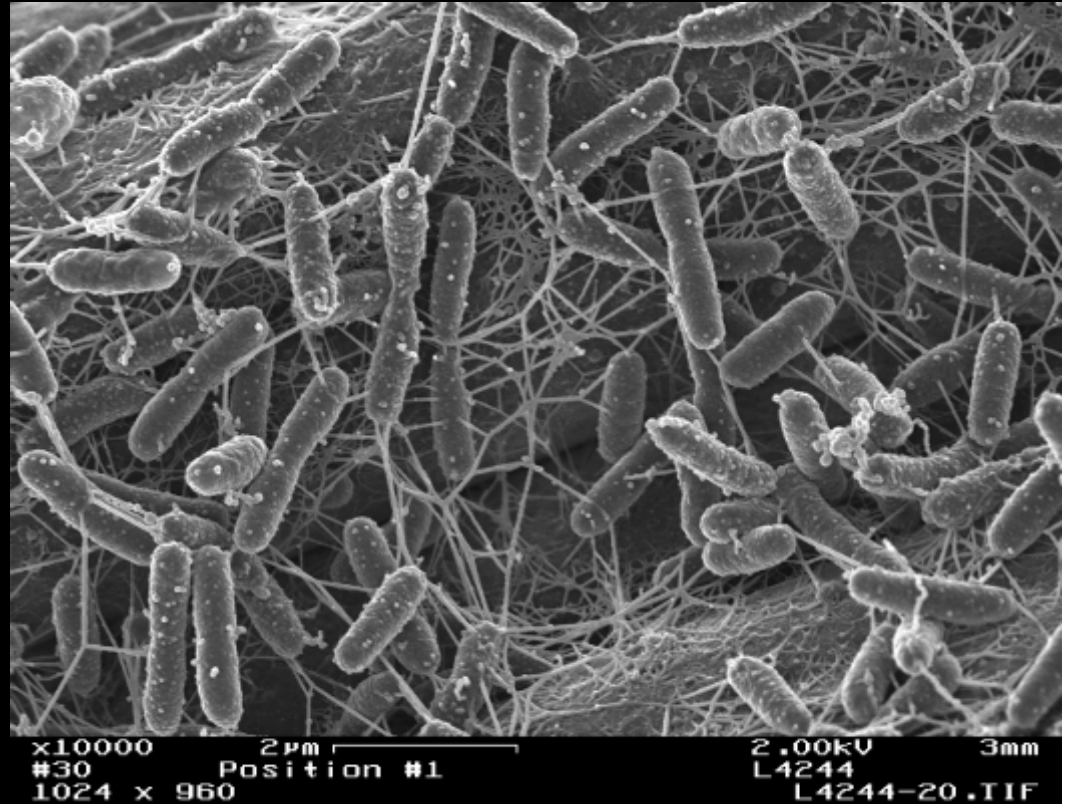


# New finding: bacteria use “nanowires”



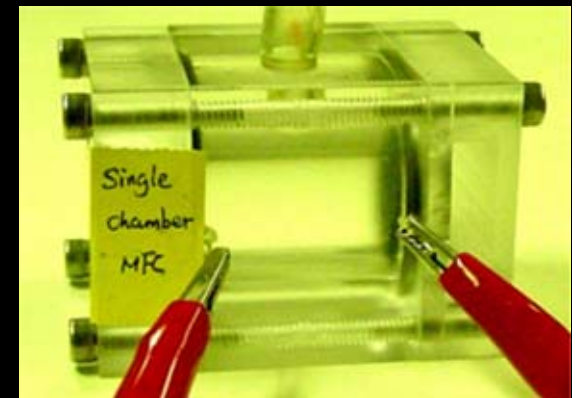
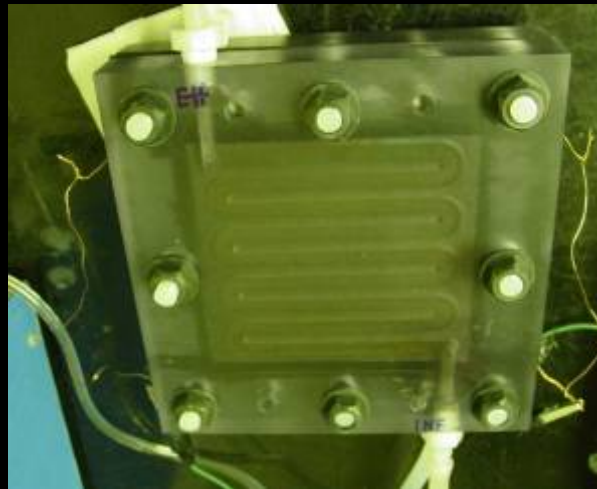
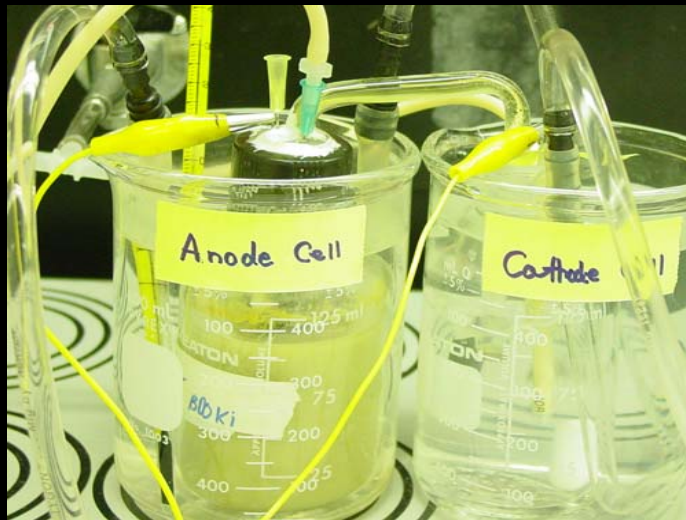
Bacteria that can transfer electrons directly to the electrode:

- *Geobacter sulfurreducens*
- *Alteromonas* sp.
- *Shewanella* spp.



# Current level of development:

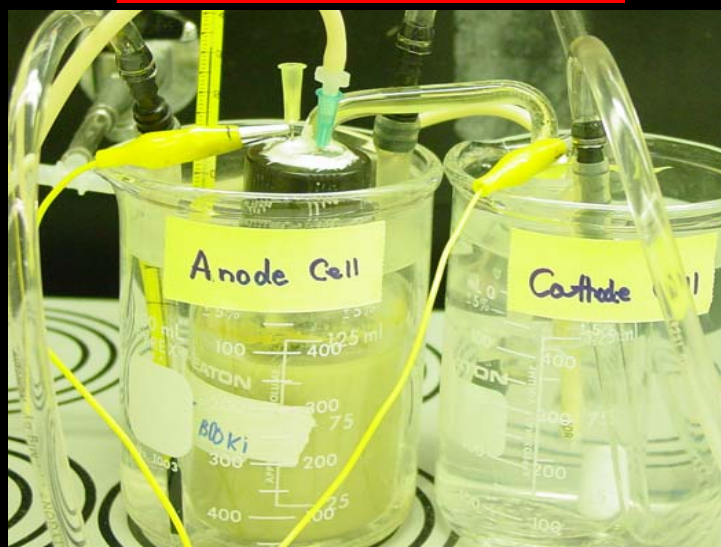
## System architecture (not microbiology) limits power





# Power density is limited by internal (system) resistance

Power: 0.3-3 mW/m<sup>2</sup>



System resistance: 66,920  $\Omega$

$$P = \frac{OCV^2 R_{ex}}{A(R_{int} + R_{ex})^2}$$

Power: 40 mW/m<sup>2</sup>



System resistance: 1,756  $\Omega$

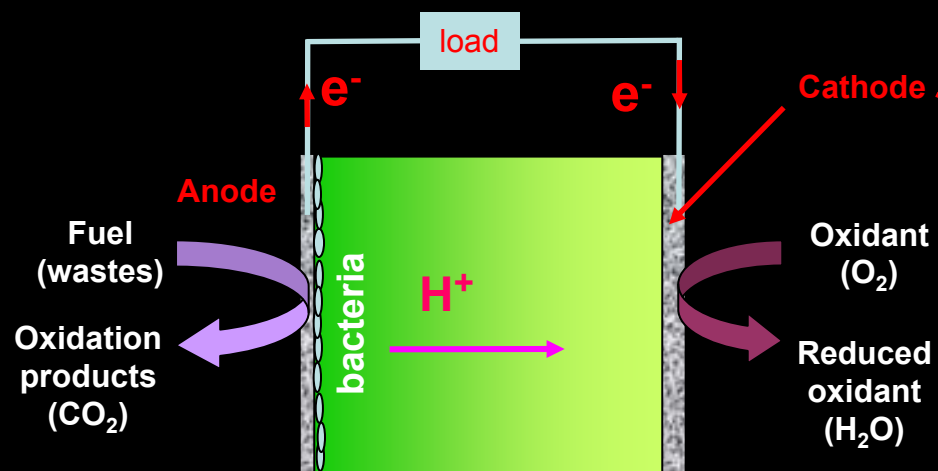
P=	Power normalized surface area
OCV=	Open circuit voltage
R <sub>ex</sub> =	External resistance
R <sub>in</sub> =	Internal resistance
A=	Electrode projected surface area



Source: Min et al., *Wat. Res* (2005)

# Air Cathode MFC

- Bacteria oxidize organic matter
- Generate electricity from any form of biodegradable organic matter



# Power production (early studies)



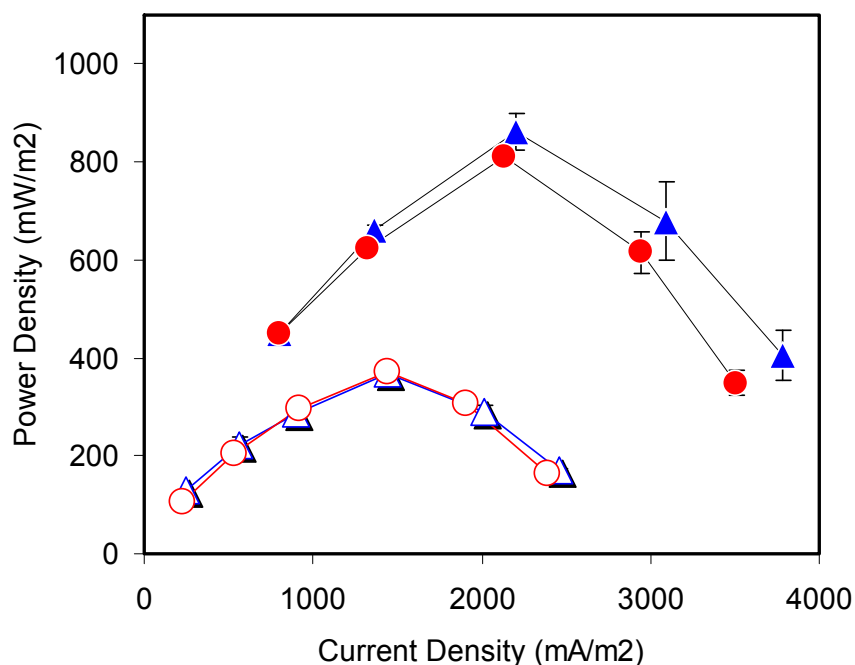
Substrate	Power (mW/m <sup>2</sup> )
Glucose	494
Acetate	506
Butyrate	309
Protein	269
Domestic wastewater	146



# Electricity from corn stover hydrolysates

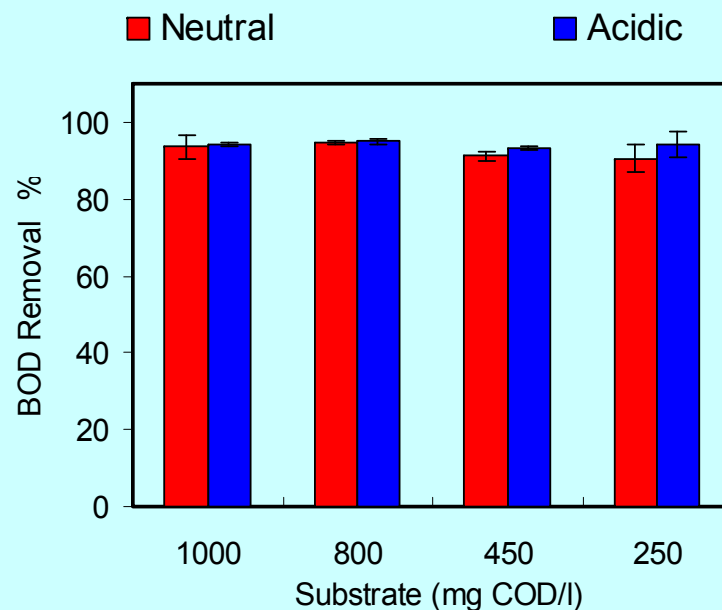
**Corn stover:** 90% of 250 million tons remains unused in fields (largest source of solid waste biomass in USA)

—△— Acidic Old Cathode      —△— Acidic New Cathode  
—○— Neutral Old Cathode      —●— Neutral New Cathode



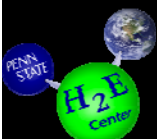
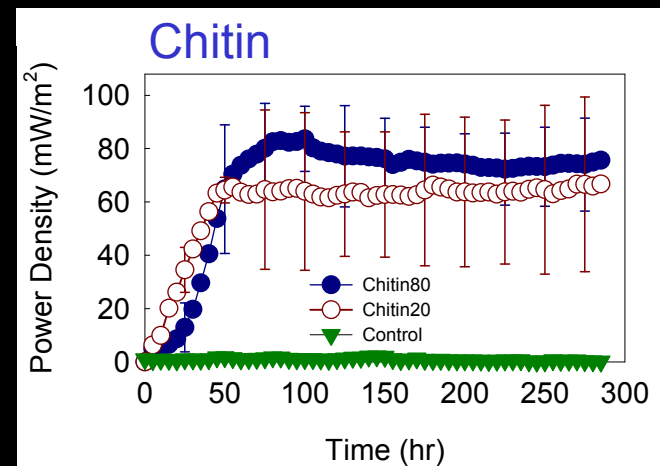
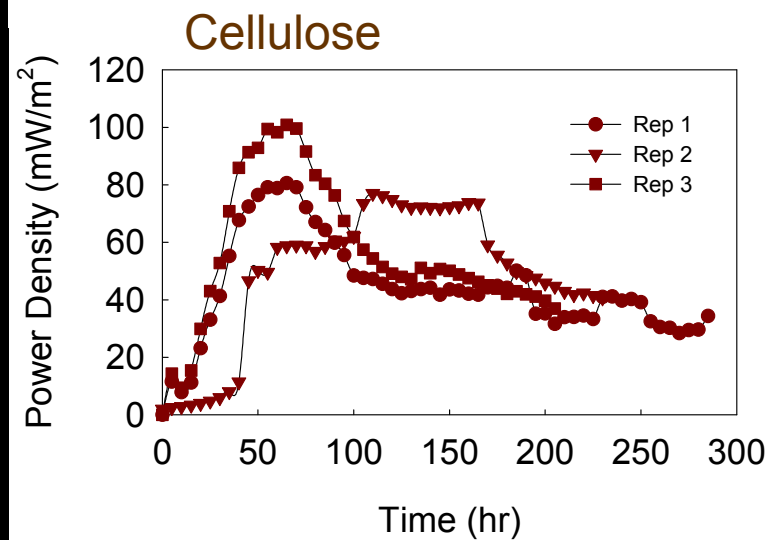
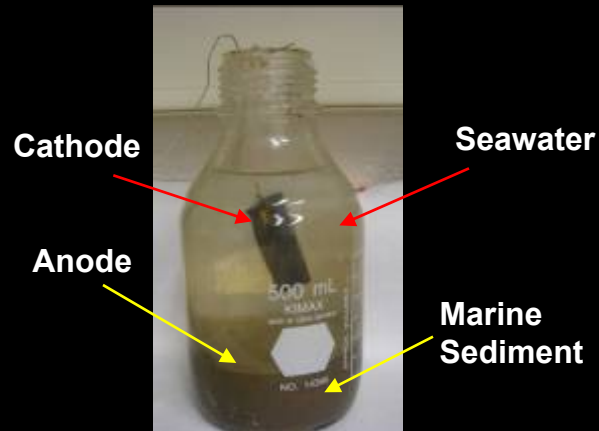
Maximum Power = 860 mW/m²

~93% removal of biodegradable organic matter



Source: Zuo et al. (2006) Energy & Fuels

# Electricity from cellulose and chitin in a sediment MFC

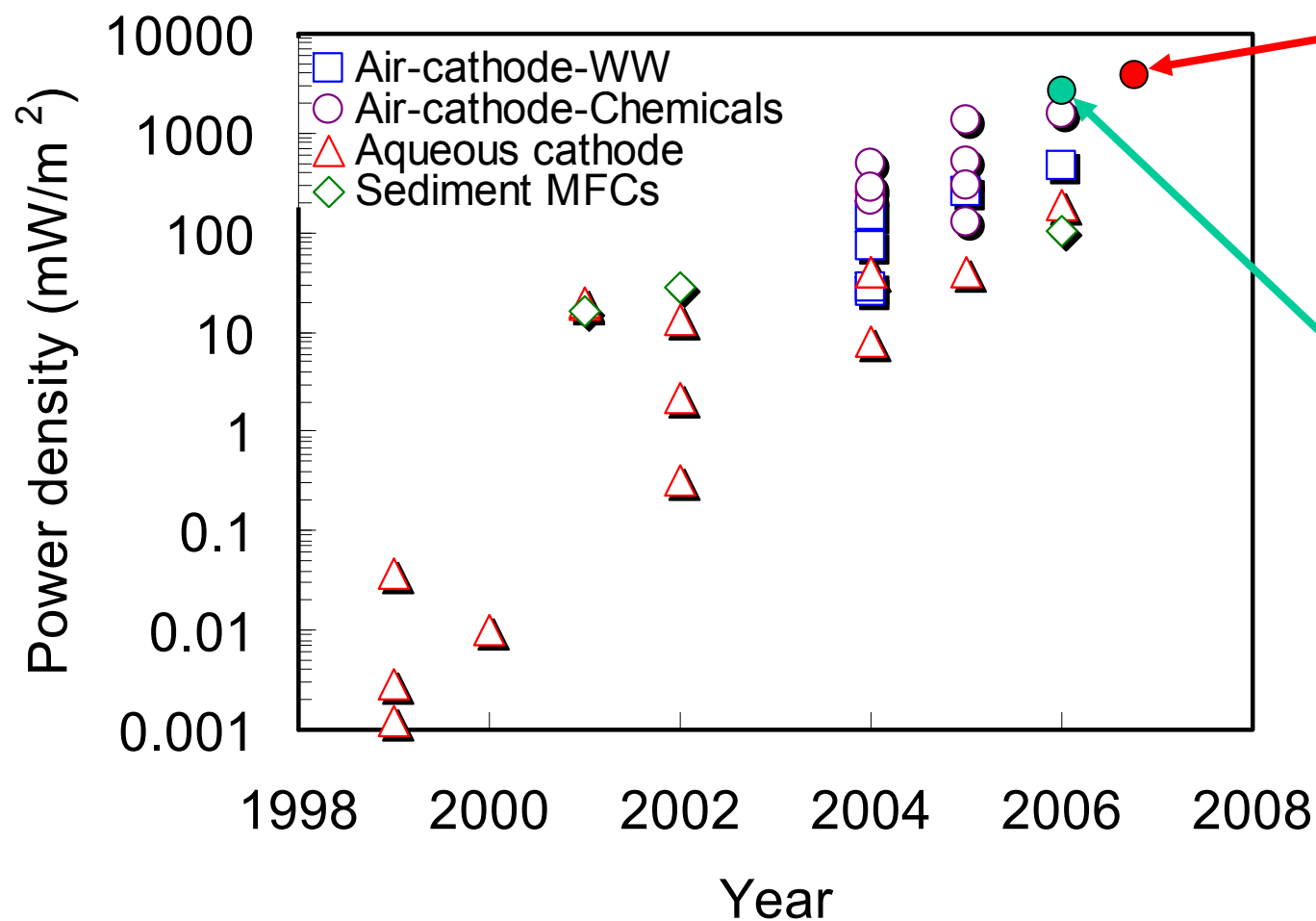


# Advances in operating conditions and materials selection

- Factors affecting performance
  - Solution conductivity
  - Electrode spacing
  - Continuous flow
- Cathode materials
  - Non-precious metal cathode catalyst
  - Diffusion layers for water control
- Anode materials
  - Treatment technique for rapid power generation (high-temperature ammonia gas)



# Power production in MFCs is improving



Start of 2007:  
2400 mW/m<sup>2</sup>  
76 W/m<sup>3</sup>

End of 2006:  
1970 mW/m<sup>2</sup>  
115 W/m<sup>3</sup>

# System Scale (Wastewater Treatment)

What will a large scale MFC system of the future look like?



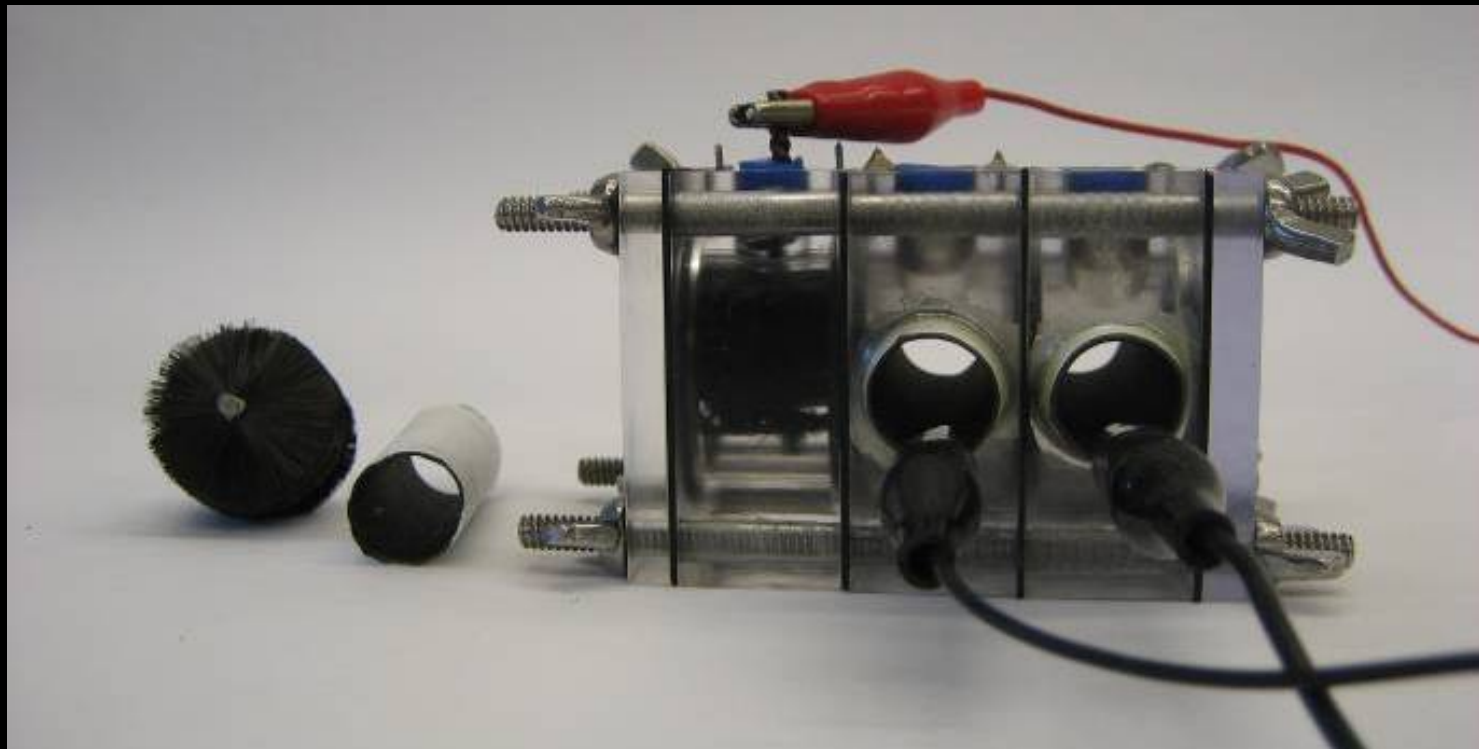
# System Scale up

- Scale up covered by two Penn State patents
  - “Materials and configuration for scalable microbial fuel cells.” Provisional patent.
  - “A bioelectrochemically assisted microbial reactor (BEAMR) that generates hydrogen gas.” (60/588,022)



# System Scale up

Brush anodes and tube  
cathode reactors



# Energy Production using MFC technologies

- Electricity production using microbial fuel cells (MFCs)
- H<sub>2</sub> Production from biomass using the MEC/BEAMR process: overcoming the “fermentation barrier”
- A path to renewable energy

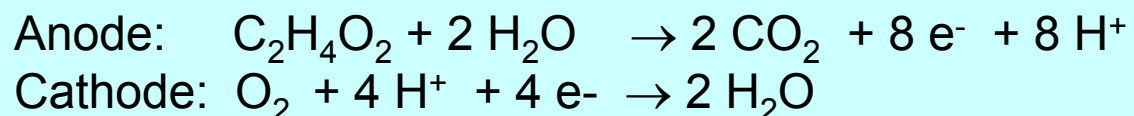




# Essentials of the MEC Process

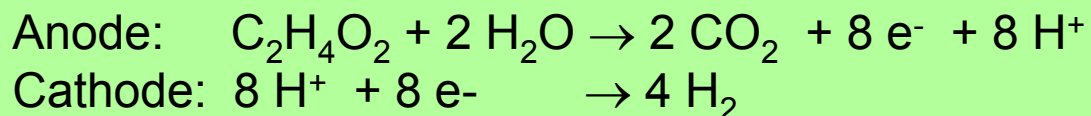
- Conventional MFC: oxygen at the cathode

- Anode potential= -300 mV
- Cathode Potential= +200 mV (+804 mV theory)
- Circuit working voltage= 200 - (-300) = 500 mV

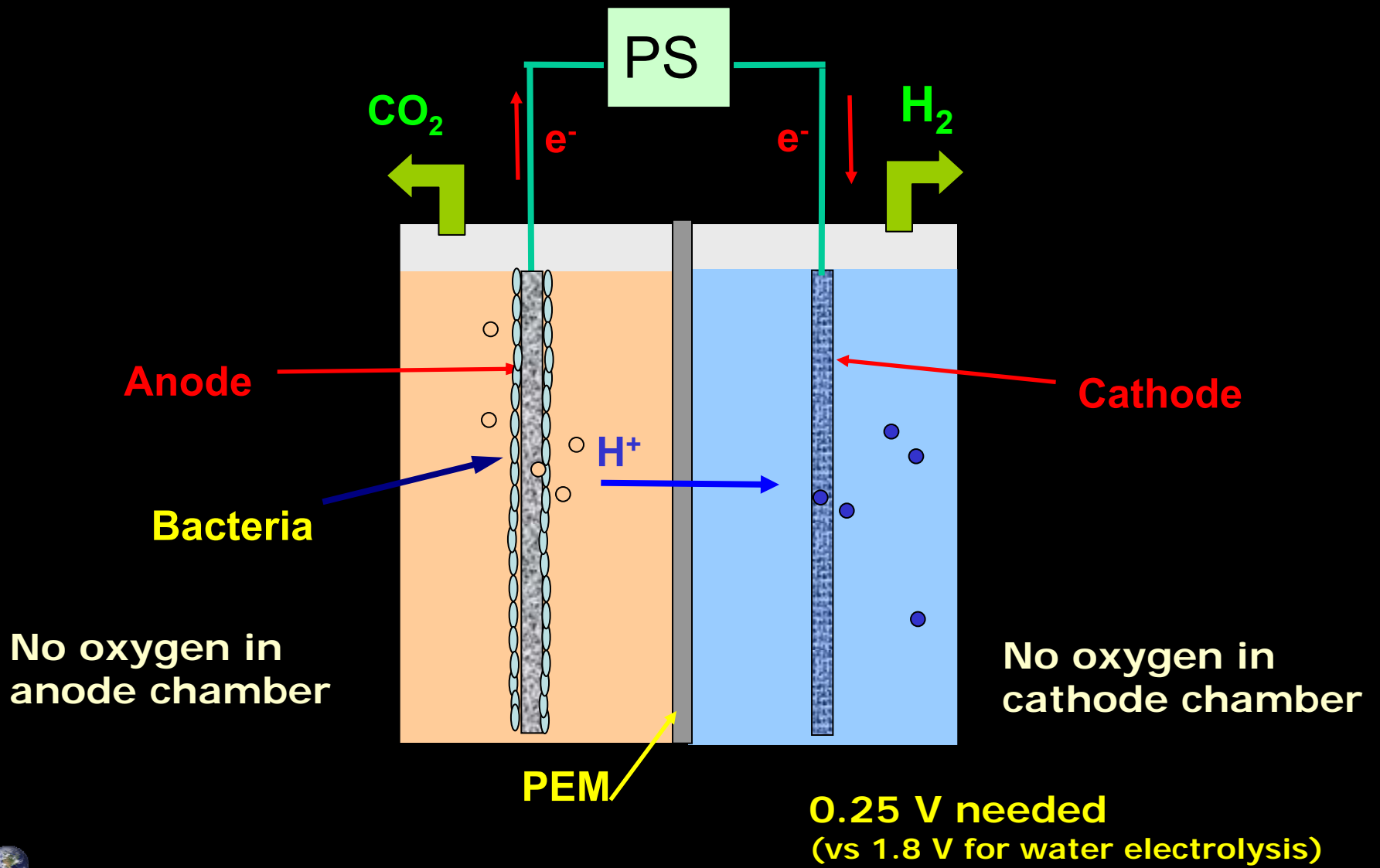


- MEC: (no oxygen)

- Anode potential= -300 mV
- Cathode potential: 0 mV
- Needed to make  $\text{H}_2$ = 410 mV (theory)
- Circuit (~300 mV) augmented with >110 mV= >410 mV



## Biomass $H_2$ - from any biomass using MECs

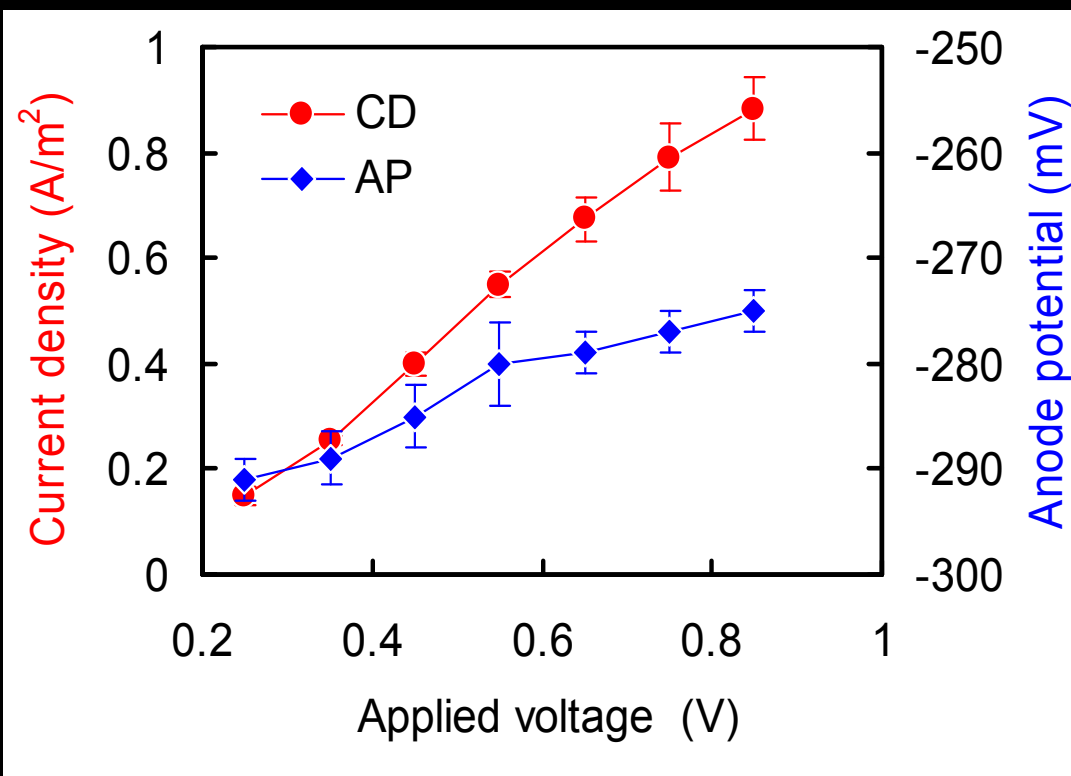


# Low applied voltages used for H<sub>2</sub> production

Minimum voltage  
needed is  $>0.25$  V  
( $=0.11$  V theory)



BEAMR voltage much  
less than that needed for  
water electrolysis (1.8 V)



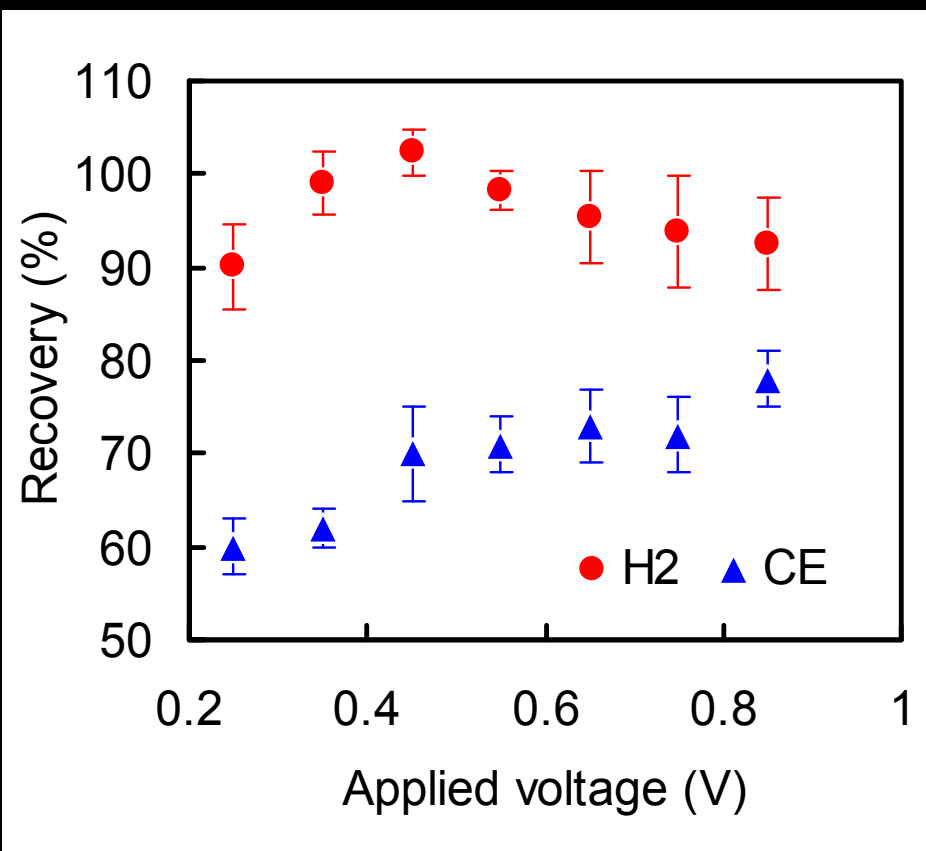
# H<sub>2</sub> Recovery

$R_{CE} = 60\%-78\%$   
(electrons from substrate)

$R_{Cat} = 90-100\%$   
(H<sub>2</sub> from electrons)

**-Overall:**

$R_{H_2} = >70\%$  ( $>0.5$  V)  
(=2.9 mol-H<sub>2</sub>/mol-acetate  
vs maximum of 4 mol/mol)



# Breakthrough in H<sub>2</sub> production rates and energy efficiencies

## Sustainable and efficient biohydrogen production via electrohydrogenesis

Shaoran Cheng and Bruce E. Logan\*

Department of Civil and Environmental Engineering, Pennsylvania State University, University Park, PA 16802

Edited by James M. Tiedje, Michigan State University, East Lansing, MI, and approved September 17, 2007 (received for review July 9, 2007)

Hydrogen gas has tremendous potential as an environmentally acceptable energy carrier for vehicles, but most hydrogen is generated from nonrenewable fossil fuels such as natural gas. Here, we show that efficient and sustainable hydrogen production is possible from any type of biodegradable organic matter by electrohydrogenesis. In this process, protons and electrons released by exoelectrogenic bacteria in specially designed reactors (based on modifying microbial fuel cells) are catalyzed to form hydrogen gas through the addition of a small voltage to the circuit. By improving the materials and reactor architecture, hydrogen gas was produced at yields of 2.01–3.95 mol/mol (50–99% of the theoretical maximum) at applied voltages of 0.2 to 0.8 V using acetic acid, a typical dead-end product of glucose or cellulose fermentation. At an applied voltage of 0.6 V, the overall energy efficiency of the process was 288% based solely on electricity applied, and 82% when the heat of combustion of acetic acid was included in the energy balance, at a gas production rate of 1.1 m<sup>3</sup> of H<sub>2</sub> per cubic meter of reactor per day. Direct high-yield hydrogen gas production was further demonstrated by using glucose, several volatile acids (acetic, butyric, lactic, propionic, and valeric), and cellulose at maximum stoichiometric yields of 54–91% and overall energy efficiencies of 64–82%. This electrohydrogenic process thus provides a highly efficient route for producing hydrogen gas from renewable and carbon-neutral biomass resources.

biofilms | cellulose | electron transport | hydrogen | microbial fuel cells

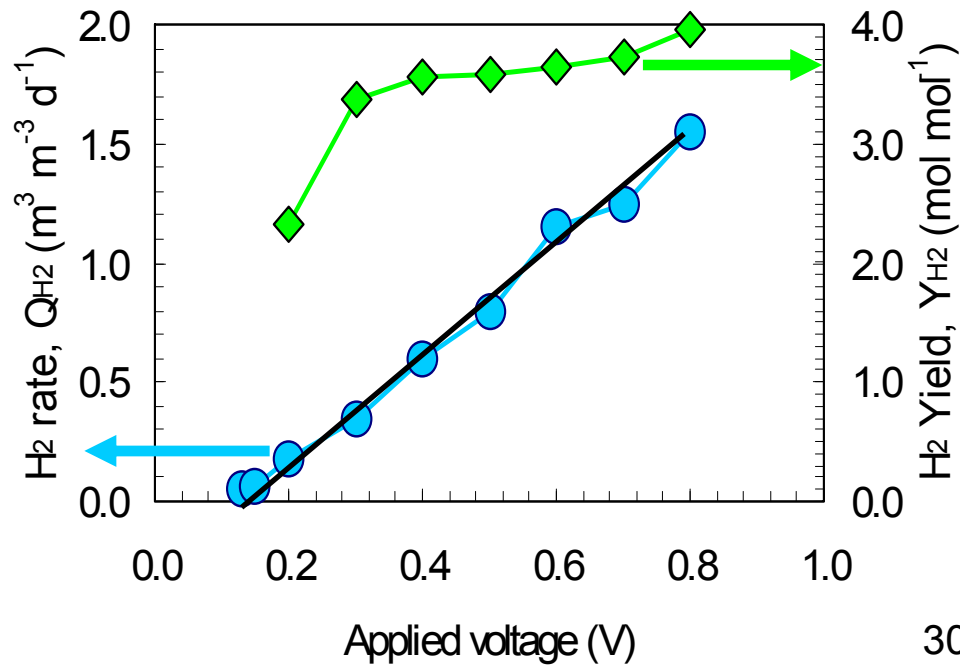
posed by the microbial formation of fermentation dead-end products, such as acetic acid. However, overall process efficiencies and hydrogen production rates have previously been low.

To substantially increase hydrogen generation rates and efficiencies, we developed a compact reactor system using chemically modified three-dimensional graphite granule anode and an anion exchange membrane. Graphite granules were treated by a high-temperature ammonia gas process to increase bacterial adhesion and overall power generation (10). Hydrogen fuel cells, and most other reactors previously examined, use a cation exchange membrane (e.g., Nafion) to keep reactor chambers separated. The membrane reduces the diffusion of hydrogen evolved at the cathode into the anode solution where it can be lost to bacterial oxidation. Cation exchange membranes preferentially transfer cations present at high concentration in solution rather than protons (e.g., Na<sup>+</sup> and K<sup>+</sup>) (11, 12), resulting in an elevated pH at the cathode that limits the hydrogen evolution reaction and a reduced pH at the anode that limits bacterial growth. Low pH cannot be used at the anode because exoelectrogenic bacteria require near-neutral pH conditions, resulting in low proton concentrations (i.e., 10<sup>-7</sup> M) which can limit the overall reaction rate. By using an anion exchange membrane, proton conduction was enhanced by protons being carried by negatively charged phosphate anions through the membrane.

### Results and Discussion

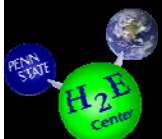
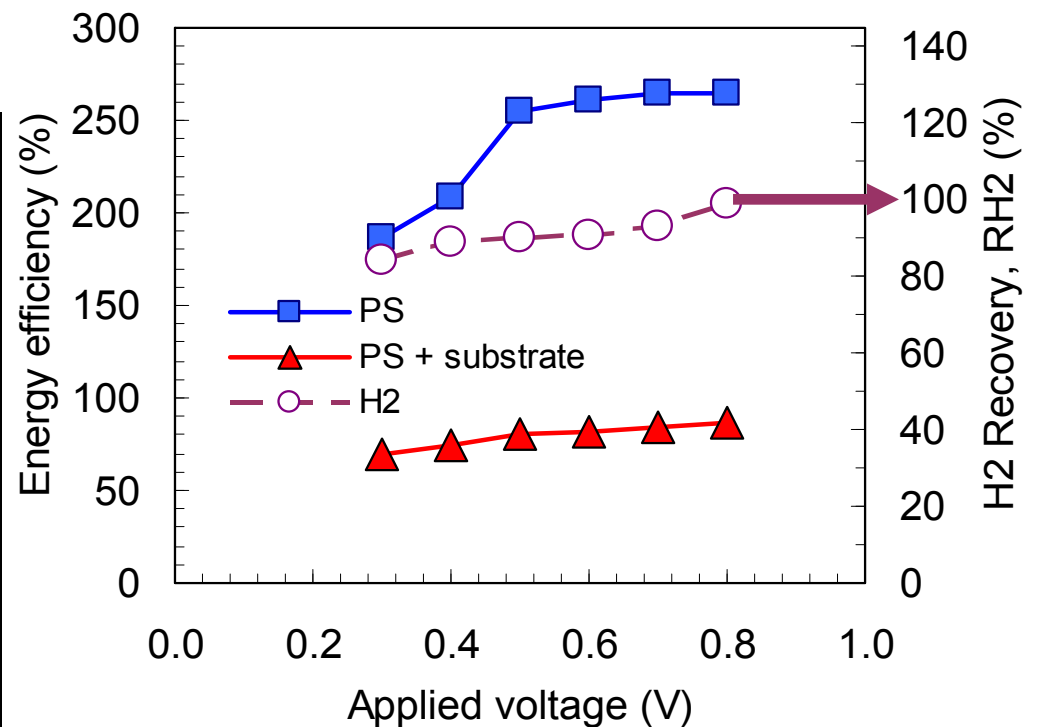
Hydrogen gas production from acetic acid, the predominant





High hydrogen yields  
except at 0.2 V

H<sub>2</sub> production increases  
with applied voltage



Source: Cheng & Logan, *PNAS* (2007)

# H<sub>2</sub> production from various substrates

Substrate	$Y_{H_2}$ (mol-H <sub>2</sub> mol-substrate <sup>-1</sup> )	$R_{H_2}$ (%)	Production rate (m <sup>3</sup> m <sup>-3</sup> d <sup>-1</sup> )	$\eta_w$ (%)	$\eta_{w+s}$ (%)
Glucose	8.55	71	1.23	266	64
Cellulose	8.20 <sup>a</sup>	68	0.11	268	63
Acetic acid	3.65	91	1.10	261	82
Butyric acid	8.01	80	0.45	285	77
Lactic acid	5.45	91	1.04	283	82
Propionic acid	6.25	89	0.72	248	79
Valeric acid	8.77	67	0.14	263	66



Source: Cheng & Logan, *PNAS* (2007)

# Energy Production using MFC technologies

- Electricity production using microbial fuel cells (MFCs)
- H<sub>2</sub> Production from biomass using the MEC/BEAMR process: overcoming the “fermentation barrier”
- A path to renewable energy





# Energy Utilization in the USA

US energy use: 97 quad

US electricity generation: 13 quad

Energy used for water infrastructure  
(water and wastewater) 0.6 quad  
(5%)

97 quad [quadrillion BTUs] = 28,400 TWh (terawatt hours)



## Energy value of wastewater

- Electricity used for water infrastructure= **0.6 quad** (~5% of all electricity)
- Energy in wastewater= **0.5 quad**
  - **0.1** quad of energy in domestic wastewater
  - **0.1** quad in food processing wastewater
  - **0.3** quad in animal wastes

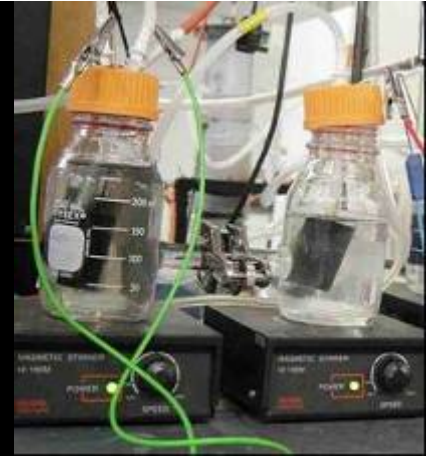
Wastewater has **9.3×** more energy than treatment consumes

(Toronto WWTP, Shizas & Bagley (2003))





# CONCLUSIONS



- MFCs represent a viable technology for simultaneous electricity generation and wastewater treatment
- The MEC/BEAMR process can overcome the “fermentation barrier” and result in high yields of hydrogen from any source of biomass
- The technology now exists for system scaleup— pilot testing is needed.

# Acknowledgements



## Current research sponsors

NSF (BES Program): 2003-2007

WERF (Busch Award): 2004

AFOSR: 2006-2009

Air Products: 2006-2008

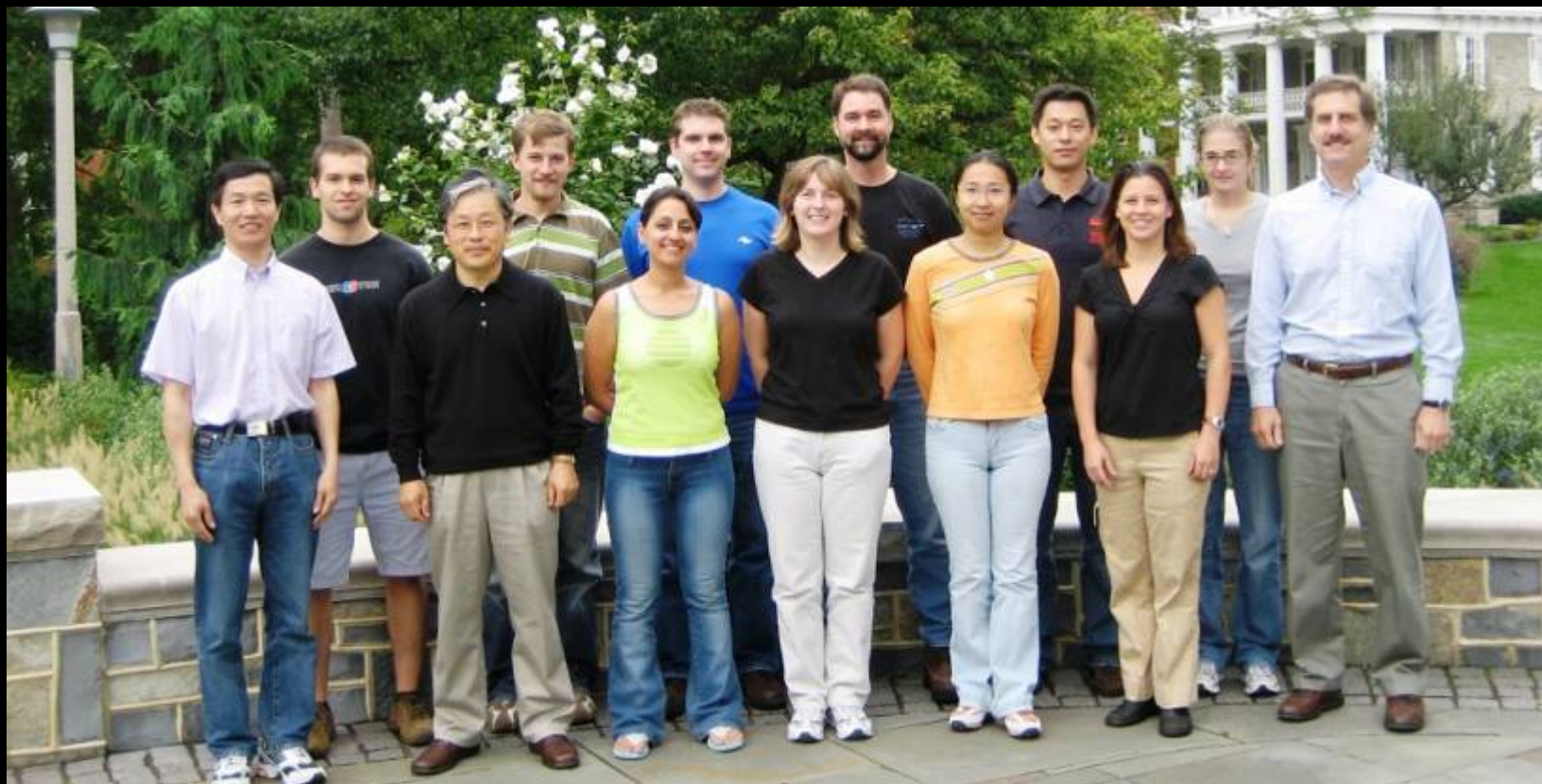
## Previous sponsors:

NSF & EPA TSE (CTS Program)

USDA-DOE



# Thanks to students and researchers in my laboratory at Penn State!

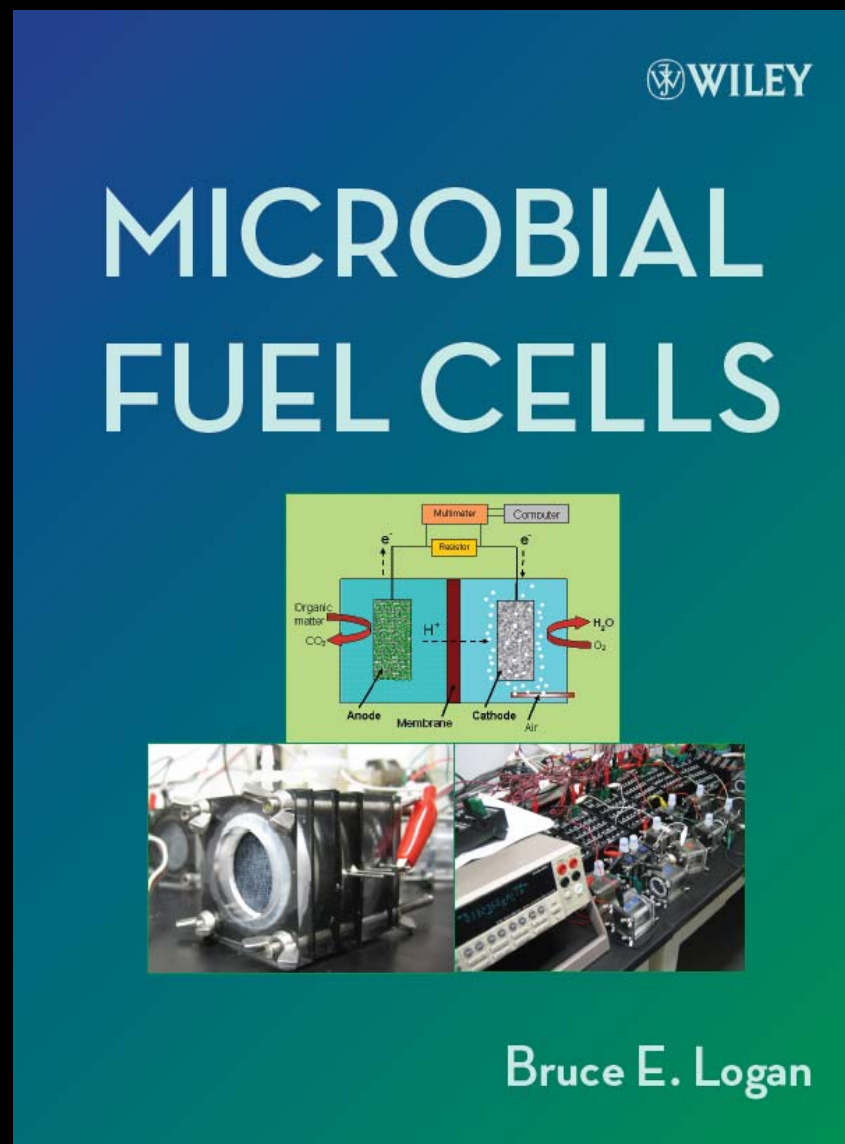


Left to right: Shaoan Cheng, Doug Call, KyeongHo Lim (visiting Prof), Markus Coenen (visitor), Farzaneh Rezai (PhD Ag&BioEng), Charles Winslow, Valerie Watson, David Jones (technician), Yi Zuo, Defeng Xing, Priscilla Selembo (PhD ChE), Rachel Wagner, [Bruce Logan]

Missing: Yujie Fang and YoungHo Ahn (visiting Profs), Jooyoun Nam (visiting student)

# Further information?

- New book on MFCs (published by John Wiley, 2008)
- Microbial Fuel Cell Symposium, May 27-29, 2008, Penn State University



# LINKS

Email: [blogan@psu.edu](mailto:blogan@psu.edu)

Web page:

[www.engr.psu.edu/ce/enve/logan.htm](http://www.engr.psu.edu/ce/enve/logan.htm)

Logan MFC webpage:

[www.engr.psu.edu/ce/enve/mfc-Logan\\_files/mfc-Logan.htm](http://www.engr.psu.edu/ce/enve/mfc-Logan_files/mfc-Logan.htm)

International MFC site:

[www.microbialfuelcell.org](http://www.microbialfuelcell.org)

MFC webcam (live video of an MFC running a fan)

[www.engr.psu.edu/mfccam](http://www.engr.psu.edu/mfccam)

