**Staining *Crithidia* with MitoTracker Red**

1. Make a 1 mM stock of MitoTracker Red CMXRos
	1. Thaw 1 vial of MitoTracker at room-temperature (keep tube in the dark by covering with foil).
	2. Resuspend MitoTracker in 94.07 μl sterile DMSO.
	3. Aliquot 10 μl per tube. Cover tubes with foil. Keep one aliquot on ice. Store remaining aliquots at -20 °C.
2. Add 20 μl MitoTracker stock to 10 ml BHI medium (2 μM final). Pre-warm medium to 27 °C (this is critical).
3. Count cells (should be mid-log phase, 2-5E7 cells/ml). Spin ~1E7 cells per sample 1000 x g 5 min.
4. Remove supernatant. Resuspend each cell pellet in 1 ml MitoTracker-BHI medium.
5. Incubate at 27 °C for 1 hour.
6. Spin at 1000 x g 5 min.
7. Resuspend in 1 ml pre-warmed BHI medium (without MitoTracker).
8. Incubate at 27 °C for 10 min.
9. Add 125 μl 36% formaldehyde to each sample (4% final).
10. Fix for 5 min at room-temperature.
11. Spin at 1000 x g 5 min.
12. Wash 2X with 1 ml 1X PBS.
13. Resuspend in 1 ml 1X PBS (1E7 cells/ml final).
14. Adhere 200 μl to poly-L-lysine-coated or charged slides for 10-20 min in humid chamber.
15. Wash slides 1X briefly and 1X 5 min in 1X PBS (in Coplin jars).
16. Permeabilize in 200 μl 0.1% Triton X-100 in PBS for 5 min in humid chamber.
17. Wash 1X briefly and 1X 5 min in PBS (Coplin jars).
18. Stain with 200 μl 0.2 μg/ml DAPI in PBS for 5 min in humid chamber.
19. Wash 1X briefly and 1X 5 min in PBS (Coplin jars).
20. Mount in 90% glycerol in PBS.