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## The Kringle of Life

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If you are a scientist and you were born the child of a scientist—even more so the child of two scientists—then it will be generally assumed that you were greatly influenced by your parents. This was certainly the case for me.

I received my undergraduate degree in Chemistry from the same Department at Carnegie Mellon University (CMU) in which my father worked. Perhaps the toughest test (literally) during my undergraduate years was having to take Physical Chemistry III: Quantum Mechanics, taught by none other than my father. More than anything, this taught me how he viewed the world and how he approached his thinking about both science and life. In his view, most things should be able to be explained mathematically from basic principles.

My father had come to CMU in 1976 from the laboratory of Kurt Wüthrich at the Eidgenössische Technische Hochschule (ETH) in Zurich where he published the first <sup>15</sup>N nuclear magnetic resonance (NMR) study on living cells [21] and explored the structure and dynamics of aluminum desferriferrichrome [24], [26, 27]. His interest in coming to CMU stemmed in large part from the Department of Chemistry's innovations in the area of NMR at that time. My father had a particular goal of applying state-of-the-art NMR technology to the structural analysis of proteins. At CMU, he pursued his studies in the company of Aksel Bothner-By, Joseph Dadok, John Pople, and others who became life-long colleagues and good friends of his.

In the mid-1970s Bothner-By and Dadok had collaborated to build the world's first 600 MHz NMR spectrometer. I remember spending Saturday mornings in my father's lab poring over the printouts of the spectra associated with the

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NMR spectrometer and using the rulers on the drafting table to find the rectilinear positions of the peaks to extrapolate the data in 3-D. Of course this is all now done by computers. My father innovated by using NMR to advance our understanding of the mechanism of blood clot dissolution, by studying the structure–function relationship of the kringle domains from tissue plasminogen activator (tPA) and other fibrinolytic proteins [2, 4, 7–8, 11, 30–32, 35–40]. tPA is now widely used to treat blood clots in thrombolytic therapy.

Many years later, one of the highlights of being a Chemistry major at CMU was that we got to use this same NMR spectrometer for our lab course. This instrument was remarkable in its appearance. It was housed in a gigantic room, and it looked as though it was built from scratch using a bunch of Radio Shack parts, combined with odds and ends from a hobby store (both of which have gone extinct). Wires ran all over the room in what seemed like a chaotic mess. Despite its unusual appearance, the NMR spectra generated by this instrument were truly spectacular!

Undoubtedly my father influenced my career in many ways, both directly and indirectly. When I was fresh out of college he helped me to secure an internship with his colleagues Ulrich Kohnert and Stephan Fisher [9–10] at a protein production facility of Boehringer Mannheim (now Roche Diagnostics) in Penzberg, Germany which had a small research arm. Although I had pursued research in yeast genetics as an undergraduate student at CMU, it was at Boehringer that I had my first exposure to industry-level research by working as a full-time technician. Although these efforts resulted in my first scientific publication [13], the experience fully convinced me that I needed to go to graduate school so that I could define and lead my own independent research interests.

After spending a wonderful nine months living in the foothills of the Bavarian Alps, I was delighted to go off to UC Berkeley to pursue my PhD. Now I was really following in the footsteps of my parents who had first arrived in the USA to study at Berkeley in 1963. Everywhere I went on campus had ties to my family history and the tumultuous 60 s and 70 s, from the Free Speech Movement to the



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anti-Vietnam War demonstrations. My father had great stories about working in the lab of Joe Neilands, who was an ardent activist during the Vietnam war. He likewise told tales of working side-by-side in the lab with Kary Mullis, whose antics and late-night experimenting were novel to my immigrant parents [33]. It was at Berkeley working with Joe Neilands and Melvin Klein that he began his lifelong work using NMR to explore solution structures of peptides with a particular focus on ferrichrome, a cyclic hexapeptide siderophore [15–20, 22, 23, 25].

Many other friends and colleagues my parents had from their time at Berkeley were still in the area, making it an inviting place for me. During my time there my mother was awarded a plaque that now hangs in Cory Hall, honoring her for being one of the first women who earned their PhDs in electrical engineering at UC Berkeley [41]. Although my mother recalls that it was odd to be in class with only men, she found the scientific environment to be highly motivating.

For my PhD thesis, I chose to work in the area of biophysics and protein folding with Prof. Susan Marqusee in Stanley Hall, the old Biochemistry and Virus Laboratory building. My doctoral work also employed NMR, which I used to measure protein stability and dynamics by hydrogen deuterium exchange in the recombinant scrapie prion protein [14] and T4 lysozyme [3, 28]. To do these studies, I worked in the famous round and open Calvin Lab, where my father had done his work on ferrichromes decades earlier. This work also led me to a series of wonderful collaborations with the lab of the renowned protein NMR spectroscopist Rick Dahlquist, who was then at the University of Oregon.

As my research interests turned away from protein biophysics during my postdoctoral years at UC San Francisco in the laboratory of Prof. Joseph DeRisi, my father was enthusiastic about my new interests in transcriptomics (further advancing the, then-new, technology of DNA microarrays) and the basic molecular underpinnings of the biology of the malaria parasite [29]. This work ultimately earned me an assistant professorship at Princeton University in 2005.

Since my father had begun his early training in medicine, he was delighted that I was focusing my research on a pathogen of major global health impact and applying modern tools of molecular biology and genetics. However, my training in chemistry continued to impact my research interests. I became fascinated by the metabolism and biochemistry of the malaria parasite, and my research group began to explore emerging methods in the area of metabolomics. Over the years, we developed approaches to quantitatively measure global intracellular metabolites using a combination of mass spectrometry and NMR spectroscopy [5, 12, 34, 1, 6]. These research directions continue to thrive in my current lab at Penn State University. In the end, not only did my father influence my work, but I physically came full circle to live within reach of my hometown of Pittsburgh, Pennsylvania,

which facilitated visiting him frequently during the last years of his life.

Anyone who knew my father knows that he was a purist and a self-proclaimed "expert" on a wide range of topics. This certitude extended beyond his passion for science to his tastes in music, food, wine, travel, and art. I would like to think that many of the things he valued have contributed to who I am today. In his work and in his daily life, my father pursued his idea of perfection. While life isn't perfect, we can remember him for seeking this ideal in himself and others, and for encouraging us to always strive to achieve our best

## References

- Allman EL, Painter HJ, Samra J, Carrasquilla M, Llinás M. "Metabolomic Profiling of the Malaria Box Reveals Antimalarial Target Pathways." Antimicrobial Agents and Chemotherapy. 2016; pii: AAC.01224–16.
- Byeon IJ, Kelley RF, Llinás M (1989) 1H NMR structural characterization of a recombinant kringle 2 domain from human tissue-type plasminogen activator. Biochemistry 28(24):9350–9360. https://doi.org/10.1021/bi00450a016
- Cellitti J, Llinás M, Echols N, Shank EA, Gillespie B, Kwon E, Crowder SM, Dahlquist FW, Alber T, Marqusee S (2007) Exploring subdomain cooperativity in T4 lysozyme I: structural and energetic studies of a circular permutant and protein fragment. Protein Sci 16(5):842–851
- Christen MT, Frank P, Schaller J, Llinás M (2010) Human plasminogen kringle 3: solution structure, functional insights, phylogenetic landscape. Biochemistry 49(33):7131–7150. https://doi.org/10.1021/bi100687f
- Cobbold SA, Vaughan AM, Lewis IA, Painter HJ, Camargo N, Perlman DH, Fishbaugher M, Healer J, Cowman AF, Kappe SH, Llinás M (2013) kinetic flux profiling elucidates two independent acetyl-CoA biosynthetic pathways in plasmodium falciparum. J Biol Chem 288(51):36338–36350
- Cowell AN et al (2018) Mapping the malaria parasite druggable genome by using in vitro evolution and chemogenomics. Science. https://doi.org/10.1126/science.aan4472
- De Marco A, Hochschwender SM, Laursen RA, Llinás M (1982) Human plasminogen. Proton NMR studies on kringle 1. J Biol Chem 257(21):12716–12721
- Gehrmann M, Briknarová K, Bányai L, Patthy L, Llinás M (2002)
   The col-1 module of human matrix metalloproteinase-2 (MMP-2): structural/functional relatedness between gelatin-binding fibronectin type II modules and lysine-binding kringle domains.

   Biol Chem 383(1):137–148. https://doi.org/10.1515/BC.2002.014
- Hu CK, Kohnert U, Wilhelm O, Fischer S, Llinás M (1994) Tissuetype plasminogen activator domain-deletion mutant BM 06.022: modular stability, inhibitor binding, and activation cleavage. Biochemistry 33(39):11760–11766. https://doi.org/10.1021/bi002 05a011
- Hu CK, Kohnert U, Sturzebecher J, Fischer S, Llinas M (1996)
   Complexation of the tissue plasminogen activator protease with benzamidine-type inhibitors: interference by the kringle 2 module. Biochemistry 35(10):3270–3276. https://doi.org/10.1021/bi9515026
- Ji WR, Castellino FJ, Chang Y, Deford ME, Gray H, Villarreal X, Kondri ME, Marti DN, Llinás M, Schaller J, Kramer RA, Trail PA (1998) Characterization of kringle domains of angiostatin as



- antagonists of endothelial cell migration, an important process in angiogenesis. FASEB J 12(15):1731–1738. https://doi.org/10.1096/fasebj.12.15.1731
- Ke H, Lewis IA, Morrisey JM, McLean KJ, Ganesan SM, Painter HJ, Mather MW, Jacobs-Lorena M, Llinás M, Vaidya AB (2015) Genetic investigation of tricarboxylic acid metabolism during the Plasmodium falciparum life cycle. Cell Rep 11(1):164–174. https:// doi.org/10.1016/j.celrep.2015.03.011 (Epub 2015 Apr 2)
- Kohnert U, Wozny M, Llinás M, Roos A, Fischer S (1995) Active site labeling with dansyl-glutamyl-glycyl-arginyl chloromethyl ketone demonstrates the full activity of the refolded and purified tissue-type plasminogen activator variant BM 06.022. Appl Biochem Biotechnol 55(2):157–166
- Liu H, Farr-Jones S, Ulyanov NB, Llinás M, Marqusee S, Groth D, Cohen FE, Prusiner SB, James TL (1999) Solution structure of Syrian hamster prion protein rPrP (90-231). Biochemistry 38(17):5362–5377
- Llinás M, Klein MP, Neilands JB (1970) Solution conformation of ferrichrome, a microbial iron transport cyclohexapeptide, as deduced by high resolution proton magnetic resonance. J Mol Biol 52(3):399–414. https://doi.org/10.1016/0022-2836(70)90409-2
- Llinás M, Klein MP, Neilands JB (1972) The solution conformation of the ferrichromes. II. Proton magnetic resonance of metal-free ferricrocin and ferrichrysin, conformational implications. Int J Pept Protein Res 4(3):157–166
- Llinás M, Klein MP, Neilands JB (1972) Solution conformation of the ferrichromes. A comparative proton magnetic resonance study of glycine- and serine-containing ferrichromes. J Mol Biol 68(2):265– 284. https://doi.org/10.1016/0022-2836(72)90213-6
- Llinás M, Klein MP, Neilands JB (1973) The solution conformation of the ferrichromes. IV. pH dependence of the individual slow amide hydrogen-deuterium exchange in alumichrome. J Biol Chem 248(3):915–923
- Llinás M, Klein MP, Neilands JB (1973) The solution conformation of the ferrichromes. V. The hydrogen exchange kinetics of ferrichrome analogues; the conformational state of the peptides. J Biol Chem 248(3):924–931
- Llinás M, Wilson DM, Neilands JB (1973) Effect of metal binding on the conformation of enterobactin. A proton and carbon-13 nuclear magnetic resonance study. Biochemistry 12(20):3836–3843. https://doi.org/10.1021/bi00744a007
- Llinás M, Wüthrich K, Schwotzer W, Von Philipsborn W (1975) 15N nuclear magnetic resonance of living cells. Nature 257(5529):817–818. https://doi.org/10.1038/257817a0
- Llinás M, Wilson DM, Klein MP, Neilands JB (1976a) 13C nuclear magnetic resonance of the errichrome peptides: structural and strain contributions to the conformational state. J Mol Biol 104(4):853– 864. https://doi.org/10.1016/0022-2836(76)90186-8
- Llinás M, Neilands JB (1976b) The structure of two alanine containing ferrichromes: sequence determination by proton magnetic resonance. Biophys Struct Mech 2(2):105–117. https://doi.org/10.1007/BF00863704
- Llinás M, Meier W, Wüthrich K (1977) A carbon-13 spin lattice relaxation study of alumichrome at 25.1 MHz and 90.5 MHz. Biochim Biophys Acta 492(1):1–11. https://doi.org/10.1016/0005-2795(77)90208-2
- Llinás M, Wilson DM, Neilands JB (1977) Peptide strain. Conformation dependence of the carbon-13 nuclear magnetic resonance chemical shifts in the ferrichromes. J Am Chem Soc 99(11):3631–3637. https://doi.org/10.1021/ja00453a020
- Llinás M, Wüthrich K (1978a) A nitrogen-15 spin-lattice relaxation study of alumichrome. Biochim Biophys Acta 532(1):29–40. https:// doi.org/10.1016/0005-2795(78)90444-0

- Llinás M, Klein MP, Wüthrich K (1978b) Amide proton spin-lattice relaxation in polypeptides. A field-dependence study of the proton and nitrogen dipolar interactions in alumichrome. Biophys J 24(3):849–862. https://doi.org/10.1016/S0006-3495(78)85424-1
- Llinás M, Gillespie B, Dahlquist FW, Marqusee S (1999) The energetics of T4 lysozyme reveal a hierarchy of conformations. Nat Struct Biol 6(11):1072–1078
- Llinás M, Bozdech Z, Pulliam BL, Wong ED, Zhu J, DeRisi JL (2003) The transcriptome of the intraerythrocytic developmental cycle of *Plasmodium falciparum*. PLoS Biol 3(12):426
- Marti DN, Schaller J, Llinás M (1999) Solution structure and dynamics of the plasminogen kringle 2-AMCHA complex: 3(1)helix in homologous domains. Biochemistry 38(48):15741–15755. https://doi.org/10.1021/bi9917378
- Marti DN, Hu CK, An SS, von Haller P, Schaller J, Llinás M (1997) Ligand preferences of kringle 2 and homologous domains of human plasminogen: canvassing weak, intermediate, and high-affinity binding sites by 1H-NMR. Biochemistry 36(39):11591–11604. https:// doi.org/10.1021/bi971316v
- Motta A, Laursen RA, Llinás M, Tulinsky A, Park CH (1987) Complete assignment of the aromatic proton magnetic resonance spectrum of the kringle 1 domain from human plasminogen: structure of the ligand-binding site. Biochemistry 26(13):3827–3836. https://doi.org/10.1021/bi00387a014
- Mullis K (1998) Dancing Naked in the mind field. Pantheon Books, New York, p 222
- Olszewski KL, Morrisey JM, Wilinski D, Burns JM, Vaidya AB, Rabinowitz JD, Llinás M (2009) Host-parasite interactions revealed by *Plasmodium falciparum* metabolomics. Cell Host Microbe 5(2):191–199
- Ozhogina OA, Grishaev A, Bominaar EL, Patthy L, Trexler M, Llinás M (2008) NMR solution structure of the neurotrypsin Kringle domain. Biochemistry 47(47):12290–12298. https://doi.org/10. 1021/bi8005557
- Petros AM, Gyenes M, Patthy L, Llinás M (1988) Analysis of the aliphatic 1H-NMR spectrum of plasminogen kringle 4. A comparative study of human, porcine, bovine and chicken homologs. Eur J Biochem 170(3):549–563. https://doi.org/10.1111/j.1432-1033. 1988.tb13734.x
- Ramesh V, Petros AM, Llinás M, Tulinsky A, Park CH (1987) Proton magnetic resonance study of lysine-binding to the kringle 4 domain of human plasminogen. The structure of the binding site. J Mol Biol 198(3):481–498. https://doi.org/10.1016/0022-2836(87) 90295-6
- Rejante MR, Byeon IJ, Llinás M (1991) Ligand specificity of human plasminogen kringle 4. Biochemistry 30(46):11081–11092. https://doi.org/10.1021/bi00110a010
- Thewes T, Ramesh V, Simplaceanu EL, Llinás M (1988) Analysis
  of the aromatic 1H-NMR spectrum of the kringle 5 domain from
  human plasminogen. Evidence for a conserved kringle fold. Eur
  J Biochem 175(2):237–249. https://doi.org/10.1111/j.1432-1033.
  1988.tb14189.x
- Thewes T, Constantine K, Byeon IJ, Llinás M (1990) Ligand interactions with the kringle 5 domain of plasminogen. A study by 1H NMR spectroscopy. J Biol Chem 265(7):3906–3915
- Weblink: https://newsletter.eecs.berkeley.edu/2017/08/alumni-spotl ight-estela-llinas/

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